



# MANUFACTURING CHEMISTS ASSOCIATION

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July 31, 1978

TO: Vinyl Chloride Technical Panel  
Vinyl Chloride Research Coordinators  
Vinyl Chloride Medical Subcommittee

Gentlemen:

As reported in Section 6.0 of the Record of the University of Louisville Meeting on June 14, 1978, Drs. Tamburro and O'Connell promised to send the Project Manager complete abstracts of all research reviews presented on all component studies. We are pleased to report that they have exceeded their promise by sending us complete transcripts of the actual research reviews covering Dr. Tamburro's "Overview of the University of Louisville Multi-Disciplinary Approach to the Study of Chemical Carcinogenesis" and the reports on the component research projects as well. Included in the enclosed collection of papers are copies of all slides which were presented during the June 14th meeting.

We trust you will find these research summaries helpful and, for the benefit of those of you who were unable to attend, they afford an excellent insight into the overall value of this research program.

Sincerely,

J. T. Seawell  
Project Manager  
Vinyl Chloride Research

JTS:ep  
Enclosure

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July 24, 1978

TO: Vinyl Chloride Technical Panel

Gentlemen:

Dr. Torkelson has requested that the enclosed reprint entitled "Comparison of the Fate of Vinyl Chloride following Single And Repeated Exposure in Rats", by P. G. Watanabe, et al., Toxicology and Applied Pharmacology 44, 391-399(1978), be sent to members of the Technical Panel.

Sincerely,



J. T. Seawell  
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Vinyl Chloride Research

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## Comparison of the Fate of Vinyl Chloride following Single And Repeated Exposure in Rats

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Comparison of the Fate of Vinyl Chloride following Single and Repeated Exposure in Rats. WATANABE, P. G., ZEMPEL, J. A., AND GEHRING, P. J. (1978). *Toxicol. Appl. Pharmacol.* 44, 391-399. Rats were exposed by inhalation to 5000 ppm of nonlabeled vinyl chloride (VC) 6 hr/day, 5 days/week for 7 weeks. On the last day of repeated exposure <sup>14</sup>C-labeled VC was used. The fate of the <sup>14</sup>C|VC was compared in a group of rats exposed repeatedly to a group exposed simultaneously for a single 6 hr period to 5000 ppm of <sup>14</sup>C|VC. The routes and rates of excretion of <sup>14</sup>C activity were the same for the two experimental groups. The activity of microsomal enzymes, as reflected by aniline hydroxylase and *p*-nitroanisole *O*-demethylase derived from 9000 g liver supernatants, was essentially the same in rats exposed once or repeatedly or in nonexposed control rats. Covalent binding to hepatic macromolecules was greater in rats repeatedly exposed as compared to those subjected to a single exposure. These results indicate that repeated exposure to VC does not induce its biotransformation. However, the increase in hepatic macromolecular binding indicates that repeated exposure augments the reaction of electrophilic metabolites with macromolecules, and this may be expected to enhance potential toxicity including carcinogenicity.

While many pharmacokinetic and metabolic studies have been conducted on vinyl chloride (VC), most of these investigations have concentrated on the fate of VC during and following single exposure. Since cancer has been induced by long-term repeated exposure of both experimental animals and man to VC, it is important to consider alterations in the disposition of VC in the body which may occur after repeated exposure.

In preliminary studies, monochloroacetic acid was tentatively identified as a major urinary metabolite following repeated exposure (6 hr/day, 5 days/week for 9 weeks) to 5000 ppm of nonlabeled VC (Hefner *et al.*, 1975). However, this metabolite has not been found in rats given a single exposure to <sup>14</sup>C|VC (Watanabe *et al.*, 1976). This raised the question of whether the biotransformation of VC may be altered upon repeated exposure.

A change in the biotransformation of VC after repeated exposure is particularly important in assessing the hazard of exposure to VC since evidence suggests that VC is metabolized to a reactive metabolite which is ultimately responsible for its toxic manifestations. This concept is supported by studies showing an enhancement of the

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mutagenic activity of VC to bacteria when microsomal and soluble enzymes are added to the system to provide for metabolic transformation (Bartsch *et al.*, 1975; Malavielle *et al.*, 1975; Rannug *et al.*, 1974). In addition, recent reports demonstrated that liver microsomal enzymes, *in vitro*, mediate the binding of VC metabolites to the microsomes (Kappus *et al.*, 1975), protein sulfhydryl groups, RNA (Bolt *et al.*, 1975), and adenosine (Barbin *et al.*, 1975). That the biotransformation of VC is involved intimately in its toxic response was further substantiated when Reynolds *et al.* (1975a) showed that acute hepatotoxicity could be produced by VC if the animals were pretreated with phenobarbital or Arochlor 1254, inducers of microsomal enzyme activity. Reynolds *et al.* (1975b) also reported that a single 6-hr exposure to 5% VC deactivates cytochrome P-450 and other components of the mixed-function oxidase system.

Since there is evidence suggesting that repeated exposure to VC may alter its biotransformation and since the metabolism of VC is associated with its toxicity, the objective of the current study was to determine whether the fate of VC is indeed altered with repeated exposure. This was accomplished by exposing rats to nonlabeled VC for 7 weeks and, on the last exposure, to  $^{14}\text{C}$ -labeled VC. The fate of the [ $^{14}\text{C}$ ]VC was then followed for 72 hr and compared to animals receiving a single exposure.

#### METHODS

**Material.** Vinyl chloride gas (Matheson Gas Products) of 99.9% minimum purity was used throughout the study.  $^{14}\text{C}$ -Labeled VC was synthesized from [ $1,2\text{-}^{14}\text{C}$ ]-dichloroethane (New England Nuclear, Lot No. 819-292, 4.8 mCi/mmol) immediately prior to use (Wagner and Muelder, 1975). The synthesized [ $^{14}\text{C}$ ]VC has been reported to be 95–96% radiochemically pure (Wagner *et al.*, 1975). Nonlabeled VC (Matheson Gas Products) was mixed with the  $^{14}\text{C}$  material to obtain the desired specific activity. Typically, 40 ml of the [ $^{14}\text{C}$ ]VC–helium gas mixture was injected into a 10-liter Saran bag (Anspec, Inc.) containing the desired quantity of nonlabeled VC.

**Animals.** Male Sprague–Dawley rats (Spartan Research Laboratory) with an initial weight of 160–180 g were used in the study. All animals were housed in rooms in which constant humidity, temperature, and 12 hr light–dark cycle (8:00 AM–8:00 PM) were maintained. Food and water were provided *ad libitum* except during the exposure. Exposures were conducted between 9:00 AM and 4:00 PM (EST).

**Exposure.** All rats were exposed under dynamic conditions to a nominal concentration of 5000 ppm of VC (treated) or room air (control) in 30-liter glass inhalation chambers. VC was metered into the chamber air flow (~6 liters/min) with a dual syringe pump. The nominal concentration of VC was determined from the ratio of the rate at which the VC gas was dispensed and the total chamber air flow. The analytical concentration of VC was monitored continuously by recirculating a fraction of the chamber atmosphere through an infrared spectrophotometer (Wilks) set at 10.6  $\mu\text{m}$ . The rats repeatedly exposed to VC were exposed 6 hr/day, 5 days/week for 7 weeks (32 exposures in 44 days). The mean analytical concentration of VC over the 7 week exposure period was  $4775 \pm 908$  (SD) ppm.

On the final day of exposure the animals were subjected to a 6 hr exposure to  $^{14}\text{C}$ -labeled VC generated in the same manner as described above. On this final day of

exposure, the chamber atmosphere was also analyzed at hourly intervals by gas chromatography, and at the same times the  $^{14}\text{C}$  activity was determined by bubbling 1 ml aliquots of the chamber atmosphere into a scintillation solution containing Conci-fluor (Mallinckrodt Chemical), 2-methoxyethanol, toluene (6:11:83) (Watanabe *et al.*, 1976). The radioactivity was determined by counting in a liquid scintillation spectrometer. The mean analytical concentration determined on the final exposure day, when the animals were exposed to  $^{14}\text{C}$ -labeled VC, was  $4600 \pm 311$  (SD) ppm. The specific activity was 50 dpm/ $\mu\text{g}$  of VC.

The inhalation chamber was operated in a laboratory fume hood to prevent contamination of the working environment. After transit through the inhalation chamber the [ $^{14}\text{C}$ ]VC was adsorbed on activated charcoal. These traps were disposed as radioactive waste according to standard regulations.

*Procedure.* Eight rats were exposed repeatedly to VC as described previously. On the last day, five additional unexposed rats and the eight exposed repeatedly were exposed to 5000 ppm of [ $^{14}\text{C}$ ]VC for 6 hr. Following this final exposure to [ $^{14}\text{C}$ ]VC, three of the eight exposed repeatedly and two of the five exposed once were placed in glass Roth-type metabolism cages for the collection of urine, feces, and expired air.

Room air was drawn through the cages at 400–500 ml/min. The exiting air was passed through a series of traps to collect the expired [ $^{14}\text{C}$ ]VC and  $^{14}\text{CO}_2$ . The air leaving the chamber was passed first through a glass tube containing about 40 g of Drierite (W. A. Hammond Drierite Co.) to remove moisture. Subsequent transit through a series of two cold finger traps containing 50 ml of toluene, 2-methoxyethanol (80:20) and a single trap containing 120 ml of 5 M ethanolamine in 2-methoxyethanol enabled the collection of [ $^{14}\text{C}$ ]VC and  $^{14}\text{CO}_2$ , respectively. The cold finger traps were immersed in 2-methoxyethanol-dry ice baths throughout the collection periods. The trap for  $\text{CO}_2$  was maintained at room temperature.

Samples of excreta were collected for 72 hr after termination of exposure and analyzed for  $^{14}\text{C}$  activity. Expired VC was collected at 0.5-hr intervals for 3 hr; the  $\text{CO}_2$  trap and urine receptacle (immersed in dry ice bath) were changed at 12-hr intervals for 72 hr; and feces were collected every 24 hr. At the termination of the study (72 hr) the animals were killed by a blow to the head, and samples of tissues (fat, kidney, liver) were collected for analysis of  $^{14}\text{C}$  activity. The remaining carcass was skinned and homogenized (50% w/v) in distilled water and analyzed for  $^{14}\text{C}$  activity. The samples of excreta and tissue were prepared for scintillation counting as described previously (Watanabe *et al.*, 1976).

Carbon-14 activity was determined by counting in a Mark II or Mark III liquid scintillation spectrometer (Searle). External standard-channels ratios were used to determine the counting efficiency. The counts per minute were converted to disintegrations per minute using a standard quench curve.

The remaining rats in the groups exposed repeatedly and singly to VC, five and three respectively, were killed by a blow to the head immediately following exposure. A piece of liver was used to prepare a 9000 g supernatant in 1.15% KCl in order to determine aniline hydroxylase (LaDu *et al.*, 1971) and *p*-nitroanisole *O*-demethylase (Kinoshita *et al.*, 1966) activity. Macromolecular binding of radioactivity to hepatic tissue was determined by the method of Jollow *et al.* (1974). This method measures non-extractable radioactivity from a trichloroacetic acid precipitate which includes protein.

lipids, and nucleic acids. The carcass was analyzed for total radioactivity as described above.

A second experiment was conducted to confirm parameters obtained for binding of radioactivity to hepatic macromolecules. The methodology was the same as described above. A group of rats was repeatedly exposed to VC [ $4821 \pm 259$  (SD) ppm] for 8 weeks. On the last day of exposure [ $^{14}\text{C}$ ]VC was used and an additional four rats (singly exposed group) were added. The mean analytical concentration of the [ $^{14}\text{C}$ ]VC on the last day of exposure was  $5065 \pm 30$  (SD) ppm and the specific activity was 34 dpm/ $\mu\text{g}$  of VC.

### RESULTS

Excretion of  $^{14}\text{C}$  activity within 72 hr after a single or repeated exposure to 5000 ppm of [ $^{14}\text{C}$ ]VC is shown in Table 1. The percentage of  $^{14}\text{C}$  activity excreted by each route as well as the total milligram equivalents of VC recovered were essentially identical for the singly and repeatedly exposed groups. The majority of  $^{14}\text{C}$  activity eliminated was expired as VC *per se*.

TABLE 1  
PERCENTAGE  $^{14}\text{C}$ -ACTIVITY ELIMINATED BY RATS DURING 72 hr FOLLOWING INHALATION EXPOSURE TO 5000 ppm OF [ $^{14}\text{C}$ ]VINYL CHLORIDE<sup>a</sup>

	Single exposure (2) <sup>b</sup>		Repeated exposure (3) <sup>b</sup>	
	(%)	(mg equiv of VC)	(%)	(mg equiv of VC)
Expired:				
As VC	$54.5 \pm 3.5$	14.00	$53.7 \pm 2.1$	12.94
As $\text{CO}_2$	$8.0 \pm 1.4$	2.05	$9.6 \pm 1.6$	2.27
Urine	$27.1 \pm 2.1$	6.93	$25.7 \pm 1.4$	6.21
Feces	$3.2 \pm 2.5$	0.80	$1.4 \pm 0.4$	0.32
Carcass and tissues	$7.3 \pm 2.5$	1.89	$9.7 \pm 1.6$	2.32
Total VC recovered		25.67		24.07

<sup>a</sup> Expressed as percentage of the total  $^{14}\text{C}$  activity recovered, mean  $\pm$  SD.

<sup>b</sup> Number of rats.

The time course for expiration of [ $^{14}\text{C}$ ]VC *per se* (Fig. 1) and urinary excretion of  $^{14}\text{C}$  activity (Fig. 2) were also essentially identical for the singly and repeatedly exposed rats. The curves were fit by linear regression analysis of the logarithmically transformed data. The estimate of the apparent first-order rate constant for expiration of VC was  $0.023 \pm 0.01$  (SD)  $\text{min}^{-1}$  which corresponds to a half-life of 30 min. The elimination of urinary  $^{14}\text{C}$  activity was biphasic. An estimate of the apparent first-order rate constant for the initial portion of the urinary excretion curve from 12 to 36 hr was  $0.155 \pm 0.02$  (SD)  $\text{hr}^{-1}$  which corresponds to a half-life of 4.47 hr. The data for the slow phase of urinary excretion were extremely variable; therefore no attempt was made to estimate the excretion rate. Less than 1% of the radioactivity excreted in the urine occurred during the slow phase.

Urinary  $^{14}\text{C}$  activity was separated by thin-layer chromatography in *n*-butanol, acetone,  $\text{H}_2\text{O}$  (50:20:30) on cellulose and in *n*-butanol, acetic acid,  $\text{H}_2\text{O}$  (80:20:20) on silica gel. The profiles of radioactivity for rats exposed repeatedly or singly were qualitatively similar and no significant radioactivity was associated with the  $R_f$  value of a standard of monochloroacetic acid.

The concentration of radioactivity detected in tissue 72 hr after exposure revealed no statistically significant difference between rats exposed once or repeatedly to VC (Table

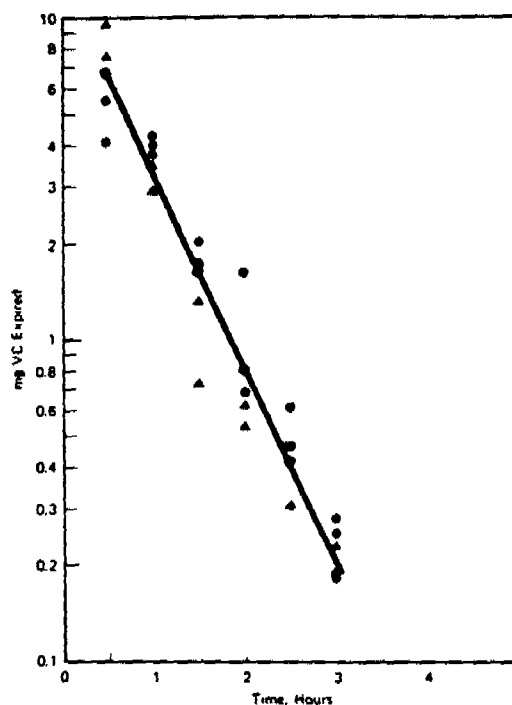


FIG. 1. Expired vinyl chloride (milligrams) versus time following a 6 hr exposure to 5000 ppm of  $^{14}\text{C}$ VC. Repeatedly exposed rats (▲) and singly exposed rats (●).

2). It does appear that, in those exposed repeatedly, more radioactivity may have been retained in the liver and skin; however, the number of animals used does not provide for an adequate statistical evaluation.

The effect of VC on xenobiotic drug metabolism by liver 9000 g supernatants, as reflected by aniline hydroxylase and *p*-nitroanisole *O*-demethylase activity, is presented in Table 3. Neither single nor repeated exposure to 5000 ppm of VC altered discernibly the enzyme activity in either system when compared to air-exposed controls.

The total amount of VC biotransformed and the hepatic macromolecular binding of  $^{14}\text{C}$  activity following single and repeated exposure are shown in Table 4. The total amount of VC biotransformed was not significantly different between the two groups. However the hepatic macromolecular binding in the first experiment appeared to be increased in the rats exposed repeatedly. When the protein binding was corrected for the

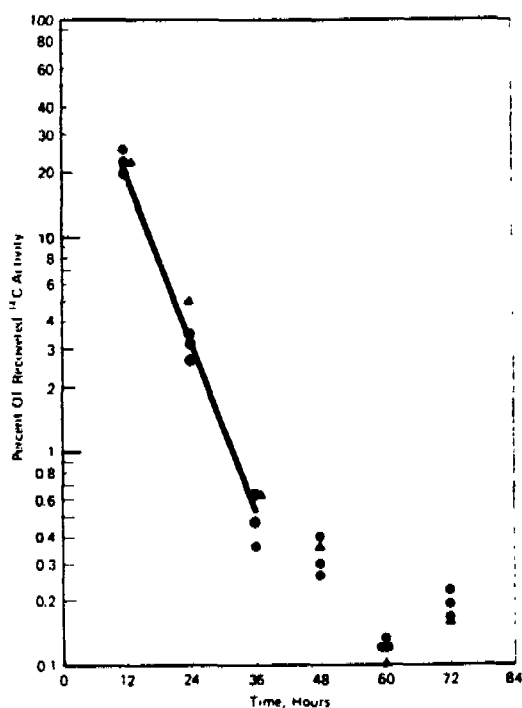


FIG. 2. <sup>14</sup>C-Activity excreted in the urine expressed as percentage of the recovered radioactivity versus time following a 6 hr exposure to 5000 ppm of [<sup>14</sup>C]VC. Repeatedly exposed rats (▲) and singly exposed rats (●).

amount of VC biotransformed ( $B/A \times 100$ ) a statistically significant increase was found. Because the increased binding in repeatedly exposed rats was not definitive in the first experiment, the experiment was repeated and the results are shown in the lower portion of Table 4. The second study confirmed the observation that rats repeatedly

TABLE 2  
PERCENTAGE <sup>14</sup>C-ACTIVITY 72 hr FOLLOWING INHALATION  
EXPOSURE OF RATS TO 5000 ppm OF VINYL CHLORIDE<sup>a</sup>

Tissue	<sup>14</sup> C Activity/g of tissue (%)	
	Single exposure (3) <sup>b</sup>	Repeated exposure (2) <sup>b</sup>
Liver	0.119 ± 0.022	0.157 ± 0.028
Kidney	0.062 ± 0.026	0.070 ± 0.006
Fat	ND <sup>c</sup>	ND <sup>c</sup>
Skin	0.046 ± 0.015	0.080 ± 0.019
Carcass	0.030 ± 0.014	0.039 ± 0.011

<sup>a</sup> Expressed as percentage of the total <sup>14</sup>C activity metabolized, mean ± SD.

<sup>b</sup> Number of rats per group.

<sup>c</sup> Not detectable, detection limit of 3 μg of VC equiv/g of fat or 0.03% <sup>14</sup>C activity metabolized/g of tissue.

exposed to VC bind about 20–25% more reactive metabolite to hepatic macromolecules than do rats exposed once. It is not clear why the magnitude of the binding was slightly greater in the second experiment. Nonetheless, these results indicate that a larger fraction of the biotransformed VC reacts covalently with hepatic macromolecules in rats exposed repeatedly.

TABLE 3  
EFFECT OF VINYL CHLORIDE ON DRUG METABOLISM BY A 9000 g  
SUPERNATANT FRACTION OF LIVER OF RATS<sup>a</sup>

	Amount ( $\mu\text{g}$ of product/g of liver/hr)	
	Aniline hydroxylase	<i>p</i> -Nitroanisole <i>O</i> -demethylase
Control (4) <sup>b</sup>	65 $\pm$ 16	226 $\pm$ 22
Single VC exposure (3) <sup>b</sup>	71 $\pm$ 7	254 $\pm$ 45
Repeated VC exposure (5) <sup>b</sup>	83 $\pm$ 10	217 $\pm$ 31

<sup>a</sup> Animals were killed immediately following the last exposure and enzyme activity was assayed. Values are mean  $\pm$  SD.

<sup>b</sup> Number of rats per group.

TABLE 4  
TOTAL METABOLISM AND HEPATIC MACROMOLECULAR BINDING FOLLOWING SINGLE OR  
REPEATED EXPOSURE OF RATS TO 5000 ppm OF VINYL CHLORIDE<sup>a</sup>

	A VC equivalents metabolized ( $\mu\text{g}$ )	B VC equivalents bound ( $\mu\text{g}/\text{g}$ of protein)	Binding corrected for metabolism (B/A $\times$ 100) <sup>b</sup>
Single exposure	9265 $\pm$ 1467	114 $\pm$ 10	1.12 $\pm$ 0.13
Repeated exposure	8718 $\pm$ 895	124 $\pm$ 10	1.43 $\pm$ 0.16 <sup>c</sup>
Experiment was repeated			
Single exposure	8746 $\pm$ 882	148 $\pm$ 25	1.69 $\pm$ 0.28
Repeated exposure	9421 $\pm$ 482	195 $\pm$ 24 <sup>c</sup>	2.07 $\pm$ 0.25 <sup>c</sup>

<sup>a</sup> Values are the mean  $\pm$  SD (three to five rats per group).

<sup>b</sup> The ratio of B/A  $\times$  100 was calculated from individual animal data.

<sup>c</sup> Statistically significantly different from the single exposure. Student *t* test ( $p < 0.05$ ).

## DISCUSSION

Repeated exposure of rats to 5000 ppm of [<sup>14</sup>C]VC did not alter discernibly the routes or rates of excretion of radioactivity or qualitatively the excretory products formed from VC or VC *per se*. These results do not substantiate the previous preliminary observation that monochloroacetate may be a major biotransformation product of VC (Hefner *et al.*, 1975).

A most significant finding in the study was a significantly increased amount of radioactivity bound covalently to macromolecules of rats exposed repeatedly to VC versus those exposed once. An associated observation was the retention of an apparently greater level of radioactivity in the liver of repeatedly exposed rats 72 hr after exposure than those exposed once. These results indicate that toxic manifestations, including carcinogenicity, associated with the reaction of reactive metabolites of VC with macromolecules may be enhanced by repeated exposure to VC.

While the binding of reactive metabolites of VC to hepatic macromolecules was enhanced following repeated exposure, no differences were observed in the activity of hepatic microsomal enzymes to the substrates aniline or *p*-nitroanisole in any of the treatment groups when compared to nonexposed control rats. Thus, it did not appear that exposure to VC at this concentration influenced microsomal metabolism. However, in contrast to these findings was the observation by Reynolds *et al.* (1975b) that the cytochrome *P*-450 content and the oxidative *N*-demethylation of aminoantipyrine and ethylmorphine were markedly depressed in rats following exposure to 50,000 ppm of VC for 6 hr. The difference between our study and that of Reynolds *et al.* (1975b) is that we used substrates which cause a "type II" binding spectra compared to their use of substrates producing a "type I" binding spectra with hepatic microsomes. In addition Reynolds *et al.* (1975b) used a 10-fold greater concentration of VC. The apparent discrepancy can be explained by the recent observation that VC causes a "type I" binding spectra when incubated with hepatic microsomes (Salmon, 1976; Ivanetich *et al.*, 1977), and furthermore it has been demonstrated (Ivanetich *et al.*, 1977) that high concentrations of VC can degrade cytochrome *P*-450. Thus it appears that VC is metabolized by the hepatic microsomal enzymes and is capable of inhibiting metabolism of other substrates by competitive inhibition or by degradation of cytochrome *P*-450.

In conclusion, the results of these studies showed that repeated exposure to high levels of VC did not alter discernibly the routes or rates of excretion of radioactivity when compared to rats subjected to a single 6 hr exposure to 5000 ppm of [<sup>14</sup>C]VC. Of particular significance was evidence that the binding of reactive metabolites of VC with hepatic macromolecules was enhanced by repeated exposure to high levels of VC. Associated with this may be expected an enhanced toxicity, including carcinogenicity. The reason for the enhanced covalent binding with repeated exposure is under investigation.

#### ACKNOWLEDGMENT

The authors wish to express their appreciation to M. M. Schlachter for technical assistance.

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